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Author: Egor Y. Plotnikov Denis N. Silachev Vasily A. Popkov Ljubava D. Zorova Irina B. Pevzner Savva D. Zorov Stanislovas S. Jankauskas Valentina A. Babenko Gennady T. Sukhikh Dmitry B. Zorov



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### Intercellular Signalling Cross-Talk: To Kill, To Heal and To Rejuvenate Short Title: Cellular communication

Egor Y. Plotnikov PhD<sup>a,d</sup>, Denis N. Silachev PhD<sup>a,d</sup>, Vasily A. Popkov<sup>b</sup>[d1], Ljubava D. Zorova PhD<sup>c,d</sup>, Irina B. Pevzner PhD<sup>a,d</sup>, Savva D. Zorov PhD<sup>b</sup>, Stanislovas S. Jankauskas PhD<sup>a</sup>, Valentina A. Babenko<sup>b,d</sup>[d2], Gennady T. Sukhikh MD<sup>d</sup>, Dmitry B. Zorov PhD<sup>a,d\*</sup>

<sup>a</sup>A.N. Belozersky Institute of Physico-Chemical Biology, Lomonosov Moscow State University, Moscow, Russian Federation

<sup>b</sup>Faculty of Bioengineering and Bioinformatics, Lomonosov Moscow State University,

Moscow, Russian Federation

<sup>c</sup>International Laser Center, Lomonosov Moscow State University, Moscow, Russian Federation

<sup>d</sup>Research Center of Obstetrics, Gynecology and Perinatology, Moscow, Russian Federation

### Abstract

Intercellular cross-talk is a fundamental process for spreading cellular signals between neighbouring and distant cells to properly regulate their metabolism, to coordinate homeostasis, adaptation and survival as a functional tissue and organ. In this review, we take a close molecular view of the underpinning molecular mechanisms of such complex intercellular communications. There are several studied forms of cell-to-cell communications considered crucial for the maintenance of multicellular organisms. The most explored is paracrine signalling which is realised through the release of diffusible signalling factors (e.g., hormones or growth factors) from a donor cell and taken up by a recipient cell. More challenging is communication which also does not require the direct contact of cells but is organised through the release of named signalling factors embedded in membranous structures. This mode of cell-to-cell communication is executed through the transfer of extracellular vesicles. Two other types of cellular cross-communication require direct contact of communicating cells. In one type, cells are connected by gap junctions which regulate permeation of chemical signals addressed to a neighbouring cell. Another type of cell communication is organised to provide a cytosolic continuum of adjacent cells joined by different tiny cell membrane extensions coined tunnelling nanotubes. In this review, we consider the various cell communication modes in the heart, and examples of processes in non-cardiac cells which may have mechanistic parallels with cardiovascular cells.

Corresponding author. A.N. Belozersky Institute of Physico-Chemical Biology, Lomonosov Moscow State University, 119992, Leninskye gory, house 1, building 40, Moscow, Russian Federation

Tel.: +7 495 9395944, fax: +7 495 9390338.

E-mail address: zorov@genebee.msu.su

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#### Introduction

Clinicians are normally occupied with the disease and ageing of their patient at systems level, however they are also aware that intercellular cross-talk is fundamental for spreading cellular signals to share information between neighbouring and distant cells to properly regulate their metabolism, according to changes in localised physical and chemical stresses, and to coordinate homeostasis, adaptation and survival as a functional tissue and organ. In this review we take a close molecular view of the underpinning molecular mechanisms of such complex intercellular communications.

There are several studied forms of cell-to-cell communications considered crucial for the maintenance of multicellular organisms, and these may have particular importance for the functional roles of stem cells in tissues. The most explored is paracrine signalling which is realised through the release of diffusible signalling factors (e.g., hormones or growth factors) from a donor cell and taken up by a recipient cell. This type of communication involves a simple release of factors into the ambient environment and subsequent interaction with a membrane receptor or channel protein. For example, the release of cytokines such as transforming growth factor  $\beta$  (TGF $\beta$ ) and growth factors such as fibroblast growth factor (FGF2) into the extracellular space by cardiac fibroblasts and myocytes which promote activation of the inflammasome, fibrosis and hypertrophy in the heart [1,2].

More challenging is communication which also does not require the direct contact of cells but is organised through the release of named signalling factors embedded in membranous structures. This mode of cell-to-cell communication is executed through the transfer of extracellular vesicles (EVs) [3], and involves formation of vesicles by a donor cell with subsequent release of vesicles into the extracellular space and absorption by a recipient cell. Such vesicles shuttle bioactive particles, proteins, lipids, metabolites and different types of nucleic acids such as DNA, mRNA, and microRNA [4-6]. Ribosomes may also be transferred using an exosomal vehicle [7,8]. In early studies, these vesicles were considered to be remnants of dead cells not playing an essential role, however subsequently all of these, some as small as 30 nm in diameter, were found to have biological roles with the potential to heal or to kill the recipient cells [9]. The size of vesicles varies from 30 nm to 1 µm. The smallest vesicles (30–100 nm) belong to the class named exosomes and larger particles (100–1000 nm) are generally named microparticles, although this classification of these vesicles by size is not strict. These two classes are different by their origin: while exosomes are formed in endosomal pathway, microparticles are the result of the cell budding. Extracellular vesicles play an

important role in the regulation of different physiological and pathological processes, thus participating in the development and progression of many diseases [9]. Extracellular vesicles, especially those produced by stem cells, cancer cells, immune cells, blood cells, and nervous system cells have become a hot study topic over recent years. The analysis of physiological fluids for EVs has become a diagnostic approach for different pathologies, including cardiovascular [10-12]. Notably, exosomes derived from stem cells can carry protective factors which can heal heart damage [13,14]. We will discuss this in detail below.

Two other types of cellular cross-communications require direct contact of communicating cells. In one type, cells are connected by gap junctions which regulate permeation of chemical signals addressed to a neighbouring cell, thus establishing electrical and mechanical synchronisation [15]. Such gap junctions can be organised by a channel with a size up to 1.5–2 nm permeable for solutes to about 1 kDa [16] including ions, oxidisable metabolites, adenine nucleotides, peptides and microRNA [17-19]. Some of these compounds (such as glucose) serve as a fuel [18], ions can regulate gating [20,21] and microRNA may be involved in a wide spectrum of activities with binary outcome. They may yield either positive effects; i.e., as shown for stem cell-derived microRNA-133a which can contribute to the activation of healing in infarct tissue [22] or similarly for microRNA-26a [23]; or they may cause negative effects; i.e., loss of miR-29 causing adverse fibrosis in the post-infarcted heart [24]. Examples involving microRNA demonstrate how the same communication transfer mechanism can provide both healing and killing functions depending on the changes of intercellular fluxes of vital components, and emphasises the regulatory importance of such a mechanism.

In contrast to gap junctions which provide an electrical continuum between cells, there is another type of cell communication which is organised to provide a cytosolic continuum of adjacent cells joined by different tiny cell membrane extensions. The first observation of organisation of the cell-to-cell channelling was made using a scanning electron microscope which showed that PC12 cells communicate by extended cellular formations coined tunnelling nanotubes (TNTs) [25]. Transmission electron microscopy showed that these extensions are organised by plasma membranes of neighbouring cells with two cytosolic contents being organised as a continuum without any structures inside which could limit exchange between cells, aside from the diameter of the nanotube itself. Sequentially, two contacting cells have not only an aqueous but also membranous continuum allowing the exchange with water soluble agents and lipid-soluble material through lateral diffusion along membranes. The transmitting chemical signal or a cargo (lipid droplets [26], vesicles or organelles [27]) are thought to be transported either passively by diffusion or as in case of a cargo transportation, by an active transport machinery. The last one often involves cytoskeletal elements such as F-actin in TNTs smaller than 100 µm in diameter and both F-actin and

microtubules in TNTs whose diameter exceeds 100  $\mu$ m [25,28,29]. Drugs which cause F-actin depolymerisation prevent formation of TNTs [30]. Tunnelling nanotubes seem to be a very secure and directed way of transporting signalling molecules between cells organised without the leak of signalling molecules into the extracellular space. The diameter of TNTs reportedly varies from 20 to 200 nm, with the length far exceeding cellular dimensions [31]. Some TNTs do not touch the substrate, thus making them highly flexible and mobile. Tunnelling nanotubes have no known unique biochemical markers but can be detected by microscopic methods such as electron (see above) and light [32] microscopy.

Lipid components have been shown to participate in the mechanism of communication between contacting cells (mostly organised by TNTs) [25,33]. It is known that in the lateral transport of molecules along the cell surface, lipid rafts play a key role in transmitting signals from receptors and EVs [34]. Participation of lipid rafts in intercellular communication is evident in mesothelioma cells contacting through TNTs that contain more lipid rafts than non-contacting cells [35].

The data on intercellular communication in the cardiovascular system are very scarce, particularly concerning the role of lipid rafts in this process. Later in more detail we will discuss the cross-talk between cardiac fibroblasts and contractile cells of the heart, which determines normal and pathological functioning of a heart [36]. In addition to this type of interaction the in vitro communication organised by TNTs between cardiac myocytes and stem cells has been shown, the importance of which lies in the possibility of stem cell differentiation into cardiomyocytes [32]. The lipid component may be one of many critical factors of cell commuting devices at least based on the fact that sites of contacts with TNTs are enriched with lipid rafts [37]. B-cells forming TNT-like cytoneme, (filipodia-like tubular extension of plasma membrane with parallel actin filaments inside the thin tube that can project to other cells conveying signalling proteins), contain a significant portion of lipid domains essential for rafts [38]. Besides the role of rafts in a cell communication organised by TNTs, the rafts are important factors in the process of uptake of EVs by a tissue including myocardium. The numerous mechanisms for EVs uptake have been documented and lipid rafts were found to be involved in both clathrin- and caveolin-mediated endocytosis as a significant part of EVs absorption by a target cell [39]. In general, lipids (particularly, cholesterol) and lipid rafts are essential components necessary for normal myocardial contractile function and ischaemic tolerance. Depletion of cholesterol aggravates both cardiac performance and cardioprotective mechanisms [40]. Later, we will describe this issue in more detail when endothelial cell - contractile cell axis will be discussed.

Thus although intercellular communication processes are likely to be highly complex due to the cellular heterogeneity of tissues and organs, the diverse modes of intercellular communication

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confers specificity, clarity of signal and compartmentalisation of signal. In the remaining sections of this review, we consider the various cell communication modes discussed above according to function, and we consider examples of processes in other cell types which may have mechanistic parallels within cardiovascular cells.

To Kill: Intercellular Communication Lessons From Tumours and the Immune System

Heart failure with preserved ejection fraction may include inflammation in its aetiology underlying its distinct structural and functional changes [41]. Pro-inflammatory cytokines such as tumour necrosis factor (TNF)- $\alpha$  and profibrotic TGF $\beta$ , are augmented in the myocardium of such patients [42]. Inflammatory cells expressing CD3, CD11a and CD45 have been detected that are associated with oxidative stress in cardiomyocytes and endothelial cells due to pro-inflammatory cytokines [43]. The subsequent transdifferentiation of cardiac fibroblasts to myofibroblasts that produces more collagen together with lower activity of metalloproteases yields fibrosis that may thus promote diastolic dysfunction in these heart failure patients [43].

This as well as other data compels us to discuss the role of communication with and among immune cells in cardiovascular pathologies.

The communication of immune with non-immune cells is organised by T-cells and antigenpresenting cells (APC). T-cell activation occurs as a result of a complex process which requires the interaction of a T-cell with major histocompatibility complex (MHC) proteins and further secretion of regulatory or cytolytic factors [44]. Analogous to cell communication in the nervous system this was called the "immunological synapse". Cell surface structures (e.g., T-cell receptor) regulating contact between the membranes of two cells are the bases of this type of communication yielding the transduction of the signal to the interior of a cell. T-cell activation consists of mostly three phases. The first phase is the T-cell polarisation when non-stimulated rounded, low motile T-cells with integrin adhesion molecules are held in an inactive state [45] after exposure to chemokines rapidly performs polarisation with formation of a front end, or lamellopodium, and a back end, or uropod [46]. The second phase is the initial adhesion organised by activated integrin triggering formation of actin-based cell protrusions enriched by T-cell receptors which form sensory contacts, subsequently immunological synapse signalling starts, and is sustained with immunological synapse maturation [47]. Interestingly, phenotypic changes in T-cells after their activation are associated with dramatic changes in mitochondrial function [48]. The pathological impact of leukocytes infiltrated in the tissue and relocated to future inflammation sites is well known. These cells cross the walls of the blood vessel starting from a layer of endothelial cells forming a first barrier for penetration [49]. The

transport of white blood cells through the endothelial barrier is a critical step for inflammatory processes driven by elevated cytokines and chemokines. Taken up from the blood stream, leucocytes first interact with receptors in the surface of endothelial cells causing leucocyte arrest and adhesion, and subsequent migration (diapedesis) across the pericyte sheath and basement membrane [50,51]. Every step of this pathway is organised by multiple factors providing cellular docking and cross-talk of leucocytes and endothelial cells. There are many reviews on mechanisms underlying this transfer (e.g., see [49]).

Exchange of molecular signals by smooth muscle cells and monocytes/macrophages may be an important step in atherogenesis. The cell dialogue between these cells results in modification of extracellular matrix composition and angiogenesis. Such communication may cause changes in the pattern of secretion of matrix proteins by smooth muscle cells which, in turn, may induce secretion by monocytes of some inflammatory angiogenic factors (such as VEGF and IL-1 $\beta$ ). This cross-talk in later stages may sequentially activate some extracellular metalloproteases and induce rupture of the plaque causing atherothrombosis [52].

Cell death triggering may be a result of concerted communication between smooth muscle cells and endothelial cells with key vasoactive players such as NO and endothelin-1 [53]. Under pathological conditions, such cellular dialogue may be altered leading to a sustained increase of vascular contractility and abnormal vascular proliferation. The communication between smooth muscle cells and endothelial cells is not limited by paracrine signalling but may also include communication via myoendothelial junctions and EVs [54,55].

Intercellular communication during carcinogenesis includes pathogenic stimulus and chronic inflammation, similar to many cardiac pathologies. The surface proteoglycan layer (glycocalyx) plays the main role in receiving primary information on the stimulus in cancer cells [56]; similarly in the heart it regulates a vascular endothelium response to physiological or pathological signals [57]. The transmission of the information involves extracellular matrix, gap junctions and other adhesion systems (reviewed in [58,59]). Cellular communications in the cancer cells environment is another example of cellular cross-talks [60].

Among numerous relations between cells which surround cancer cells, thus forming a malignant tumour, we can recall the cross-talk when one talking partner is a fibroblast. Fibroblasts are relatively undifferentiated cells with a plastic phenotype [61,62]. Apparently, their potency to be converted into different phenotypes is determined by their microenvironment. Similar to the heart, the key process of tumorogenesis is the fibroblast activation in response to tissue injury and some stimuli yielding formation of a damage-associating phenotype. In this aspect, the parallelism between tumour and heart seems obvious since while activation of fibroblasts in a tumour results in

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formation of the cancer stroma (plus inflammation), in a damaged heart it causes the cardiac remodelling as a result of fibrosis (plus inflammation) in an infarct zone and both events can be considered as deleterious for humans [63].

In a cell communication "vocabulary" which includes numerous chemical factors such as reactive oxygen species, cytokines, etc., another crucial chemical term which cells can "speak" with is mitochondrial DNA (mtDNA). It has been demonstrated that horizontal transfer of mtDNA from cell to cell may compromise respiratory function [64]. Since mtDNA is able to leave the cell [65], and this nucleic acid is known to be a component of the innate immune response, it is tantalising to suggest that mitochondrial DNA may constitute another novel mode or component of intercellular communication. Besides, transfer of mtDNA can contribute to the beneficial cardioprotective effects of mitochondrial transplantation demonstrated recently [66].

Some elements of communication through EVs and membrane lipid rafts are involved in viruses-host cell interactions. Viral infections are known to be associated with cardiac pathologies, such as myocarditis, pericarditis, and arrhythmias after infection with a dengue [67], West Nile [68] viruses and other arboviruses [69]. Viral particles may, via similar communication mechanisms to EV, interact with a host cell. For instance, HIV carries a shell made of a lipid bilayer with entrapped proteins and RNA, with a size of viral particle ranging from 100 to 120 nm. The infection with HIV is highly dependent on lipid rafts on the cellular membrane and, specifically, on the cholesterol contained in the rafts [70]. In addition, lipid rafts determine HIV internalisation and also affect the progression of the infection, particularly the release of viral particles from endosomes and permeation to the cytosol. Thus it seems tentative to suggest that lipid rafts may also play a role in the intracellular sorting of exosomes, not unlike cholesterol-sequestering agents that promote the transport of exosomes toward the apical membrane of a trophoblast facilitating their release in maternal circulation instead of equivalent process toward a fetal circulation [70]. Similar data on the role of lipid rafts in the internalisation of coxsackie virus have been reported [71].

#### To Heal: Stem Cell Interactions

As discussed above, the interaction of stem cells with other cell types may underlie potential mechanistic roles underlying cell therapy. On the one hand, the interaction of stem cells, (both exogenous and intrinsic) with specific cells within "niches" [72] may determine the fate of stem cells, the path of differentiation, proliferative potential, and, ultimately, the regenerative efficiency. Impaired perception of stem cell signals from the cell environment may lead to unpredictable consequences, including malignancy [73,74] due to formation of teratomas.

On the other hand, a growing number of recent studies address paracrine action (in the broadest meaning) of stem cells on the surrounding tissue. In this case, the signals of different origin can be transferred from stem cells to the cells of the organ, stimulating its regeneration, protecting it from damage or normalising the metabolism. In the framework of this concept, the stem cells were regarded as "cytokine factories". Indeed, it is known that they produce a significant number of biologically active molecules, such as TGF $\beta$ , VEGF, EGF, SDF-1, prostaglandin E, nitric oxide and many others [75]. These factors are released by a stem cell in the extracellular space and after binding to corresponding receptors of surrounding cells, they exert their biological effects while many of them may enter the bloodstream, causing systemic effects.

A significant portion of protective and regulatory effects of stem cells are associated with the microvesicles or exosomes released from them [76]. Such structures can contain various cytokines, and many other physiologically active components of cells, such as microRNAs, signal proteins and even organelles [14]. A great number of reviews with descriptions of the mechanics of these processes are available elsewhere including itemisation of signalling from stem cells, implemented via EVs(e.g., see [14,77-79].

Importantly, entrapment of an active compound within the vesicle solves the problem of signal dilution in the extracellular environment and it allows accurately directed delivery of signal to the targeted cells, since the surface of the exosomes can carry ligands, providing the affinity of the vesicles to specific cell types [80].

However, the greater specificity and efficiency of signal transfer between stem cells and differentiated tissue cells is provided by direct contact organised by TNTs. Tunnelling nanotubes were discovered in haematopoietic stem cells [81], between endothelial progenitor cells and cardiomyocytes [82], between mesenchymal stromal cells and cardiomyocytes [32], as well as between epithelial cells of the renal tubules and neurons [83]. To date, the structure referred to as TNTs, is described for many types of intercellular interactions, but in most cases, at least one of the partners is a stem cell [84]. Tunnelling nanotubes are a discrete cellular extension of cytoplasm, bounded by a plasma membrane which connects two cells delivering substances and signalling molecules that are transmitted through a medium or via EVs, but with far greater speed and efficiency than in the ways outlined above. However, the most intriguing result of such contact between stem and differentiated cells is the possibility that TNTs may transport cytosolic organelles, including mitochondria [25,84], which may change the metabolism of the recipient cell [66]. Evidence supports that such mitochondrial transfer within nanotubes [32,82], is distinct to each nanotube in that mitochondria can move only in one direction from cells which formed the TNT, to the cell which received the nanotube, but not vice versa [25]. In the case of stem cells, stem cells are

shown to be donors, rather than recipients of mitochondria, particularly in experimental models associated with cell damage. For example, transport of mitochondria from the multipotent mesenchymal stem cells (MMSC) into the epithelial cells of the lungs to protect them from endotoxin-induced death, maintained normal levels of ATP production and prevented lung injury in vivo [85]. Recently, the mechanism of mitochondria transport from stem cells into damaged epitheliocytes was partially resolved [86]. Finally, a recent study demonstrated that MMSC derived from induced pluripotent cells (iPS) were capable of transferring mitochondria to epithelial cells of the lungs, and could reduce the damage caused by a cigarette smoke [87]. However, although the majority of studies indicate positive effects of mitochondrial transport, sometimes donor mitochondria can have toxic effects [88]. Contact of stem and differentiated cells by TNTs has also a reciprocal effect on the stem cells. Thus, in some cases the contacts via TNTs elicited the differentiation of stem cells [32,83]. In other work, the transport of mitochondria via TNTs from smooth muscle cells have been reported to be the cause of increased proliferation of MMSC, whereas blocking the formation of TNTs abolished this effect [89]. The opposite effect was demonstrated when mature cardiomyocytes were co-cultivated with stem cells, extracted from fat or bone marrow. In this case, the transfer of mitochondria to cardiomyocytes caused their partial dedifferentiation [90]. Thus although conceptually and practically in its infancy, mitochondrial transplantation may afford rescue of cellular function [66].

### To Regulate: Communication in Heart, Brain, Vasculature and Others

In the heart, cardiac myocytes and cardiac fibroblasts are roughly in equal proportion meaning that every myocyte borders one or more fibroblasts [91]. The heart is known to frequently undergo so-called cardiac remodelling as a result of disease and ageing which is associated with structural and electrical changes in both types of cells [92,93] and strongly depends on the cells' communication. First of all, cross-talk between these cells is organised through exchange by paracrine signals (such as TNF, TGF $\beta$ , IL family, VEGF, ANG-2, endothelin-1 and others [2,94-96]). This kind of signalling can be deleterious for the heart tissue resulting in cardiac fibrosis [97]. At the same time, some excreted paracrine factors such as IL-33 and ST2 can be beneficial for the heart [98,99]. Also, paracrine factors excreted by cardiac fibroblasts can regulate electrical properties of myocytes through both direct and paracrine interaction [100-102]. One of the pathways involved in paracrine communication between cardiac myocytes and cardiac fibroblasts was suggested to involve pannexins-formed channels in the cellular membranes of these two types of cells [103,104].

It is still under debate whether cardiac myocytes and fibroblasts can communicate through gap junctions in the heart in vivo, but under in vitro co-culturing conditions they do form this kind of junction [105]. This junctional communication is reportedly deleterious, resulting in arrhythmogeneity of fibrotic myocardial cultures due to expression of connexin43 in cardiac fibroblasts [106]. Junctional coupling of myocytes and fibroblasts has been demonstrated to modulate calcium fluxes which can also contribute to incidence of arrhythmias in fibrotic heart tissue [107]. Figure 1 schematically illustrates the deleterious outcome of interaction of cardiac myocyte with cardiac fibroblast organised by electrical, biochemical and biomechanical communication, in comparison to some beneficial effects of cardiac myocyte-stem cell interactions. More detailed mechanisms of the cell-to cell communication in heart are described elsewhere [59]. Neuron-glial interactions are also important in cellular cross-talking in the developing heart and in communication between cardiac ganglia and cardiac cells of the adult heart [108]. To emphasise the importance of communication between neuronal and non-neuronal cells (basically microglia and astroglia) within the entire cellular network in CNS, the term "Neurovascular unit" has been coined [109,110]. The unit consists of the brain major cell types, namely endothelial cells, astrocytes, neurons and their axons, and other supporting cells to integrate incoming information with further release of a proper response [111].

Although in the brain, astrocytes can protect neurons from a pathological impact [112], in contrast, impaired astrocytes can release molecular factors that selectively damage neurons [113]. Interestingly, mitochondria have been reported to be involved in the cross-talk between astrocytes and neurons when neurons release impaired mitochondria with their subsequent degradation in adjacent astrocytes [114]. This implies the importance of mitochondrial transfer between neuronal and non-neuronal cells. In addition, recent work has demonstrated that neurons and astrocytes exchange with healthy mitochondria in a unidirectional way: from astrocytes to neurons [115]. This phenomenon was observed under conditions of tissue ischaemia or ischaemia-simulated conditions causing cell damage and apparently the transfer of mitochondria to these neurons fulfilled a rescuing mission in overall neuron salvage process. This result is comparable to the beneficial process of mitochondrial transplant injection to heal the damaged heart mentioned above [116,117]. There is also ultrastructural evidence for the presence of EVs containing mitochondria in the astrocytes that may, via the bloodstream, reach target organs such as the heart.

Thus, in addition to the cross-talk of neural cells by neuromediators and chemokines, neural cells can also communicate through establishing direct contacts. In these cases, gap junctions are involved by mediating the rapid diffusion and distribution of ions and transmitters to neighbouring

cells [119,120]. A principle component of gap junctions is connexin 43 (Cx43), but they also may contain Cx30, Cx26, Cx40, Cx45 and Pannexin1 providing direct contact-based cellular cross-talks [121-124].

Another important cellular partnership we observe is between endothelial cells and other cells of the tissue. A very good example for such partnership is a cross-talk between glomerular endothelial cells and podocytes and mesangial cells which is very important in aetiology of diabetic kidney disease [125]. In patients with macroalbuminuria, both podocytes damage and endothelial cells injury were observed [126]. The endothelium injury is at least partially caused by a diabetesinduced oxidative stress which activates production of heparinase, ultimately resulting in increased glomerular permeability [127]. Thus, diabetes compromises normal functioning of endothelial cells. Endothelial cells, podocytes and mesangial cells share the glomerular basement membrane on which they all sit. In normal kidney, endothelial cells transmit insulin-like growth factor (IGF) and hepatocyte growth factor (HGF) to podocytes and platelet-derived growth factor B (PDGFB) to mesangial cells, while mesangial cells send back to endothelial cells, TGF $\beta$  and integrin. In the diabetic kidney, these cross-talks are dramatically changed. The endothelial cells – podocytes vascular endothelial growth factor (VEGF) signalling, which is essential for normal kidney functioning, becomes altered [128,129]. New elements or old elements in enhanced levels, such as endothelin-1 (ET-1) [130], angiopoietins (Ang-1, Ang-2) [131] and TNF- $\alpha$  [132,133], are all implicated in renal injury. eNOS, another essential component of the renal cells cross-talk, when ablated, causes heavy albuminuria associated with podocytes injury [134]. Other factors such as prostanoids derived from activated cyclooxygenase also play a paracrine role in mediated podocytes injury [135]. Recent findings point to microRNAs (mir-143 and mir-145) as factors regulating interaction of endothelial cells with smooth muscle cells [136].

In the heart, besides earlier described communication between cardiac myocytes and fibroblasts, one of highest importance is the endothelial cell – contractile cell axis. The cardiac endothelial system is organised by a monolayer of cells covering cardiac cavities (endocardial endothelial cells) and the internal surface of the myocardial vascular system (vascular endothelial cells). Some of the factors providing communication between endocardial endothelial cells, vascular endothelial cells and cardiac myocytes are identical to those indicated for renal cellular communication. Significant attention in these links has been attracted to eNOS and its product NO, which is mostly formed by endothelial cells [137] but in some, although at much lower levels, by cardiac myocytes [138]. Expression of eNOS is modulated by numerous factors such as TGF $\beta$ , protein kinase C, TNF- $\alpha$ , HSP and others. NO is an essential factor necessary for normal cardiac functioning [139], however at high levels NO can cause pathological activation of guanylate cyclase yielding

cGMP which desensitises cardiac contractile elements to calcium ions [140]. NO activates G proteins  $(G_s \text{ and } G_i)$  stimulating Ca-channels [141]. General targets for this second messenger are proteins which can undergo nitrosylation. Critical proteins involved in excitation-contraction, such as ryanodine receptor, can be directly phosphorylated by NO resulting in myocardial contractile activation [142]. Important to this regulatory signalling are lipid rafts with their uneven distribution among planar and invaginated (caveolae) parts of the plasma membrane. Ion channel activities critical for shaping the cardiac action potential were found to be strongly dependent on the location: either being in caveolae or outside of it [143]. Importantly, there is an intracellular cross-talk between caveolae and mitochondria [144] which represents intracellular communication between the cell membrane and cellular organelles which is highly organised and proceeds with participation of G-proteins [145]. Ischaemic and pharmacologic preconditioning causes translocation of principle caveolae proteins, caveolin-1 and 3 from a cell surface to mitochondria affording a protection from ischaemia-reperfusion injury [146,147] (note, that among all caveolins caveolin-3 (Cav-3) is specific for striated muscle and certain smooth muscle cells). Caveolins regulate multiple cellular processes including cell transduction apparently through housing of numerous signalling molecules, e.g., Gprotein coupled receptors (GPCRs), thus regulating multiple associated proteins such as G<sub>i</sub>, adenylate cyclase, and effector kinases [148]. It has been found that cardiac-specific caveolin 3 expression mimics protective ischaemic preconditioning via activation of GPCR/G<sub>i</sub> signalling pathway [149]. The described picture outlines the main components participating in the communication pathway from one cell to the interior of another cell (e.g., to mitochondria with their important role in collecting survival signals [150]) to afford protection.

As we have already mentioned, depletion of cholesterol aggravates pathological changes in cardiac performance and protective stress signalling including ischaemic tolerance. Gradual depletion of sarcolemmal cholesterol content results in significant changes in myocardial function and tolerance to ischaemia/reperfusion whereas disruption of caveolae (through deletion of caveolin 3) specifically modifies ischaemic tolerance without direct effect on basic cardiac performance [40]. In parallel, ischaemic preconditioning of the heart causes translocation to mitochondria of another principal component of caveolae, connexin-43 which also affords protection not observed in inactive connexin-43 systems [151]. The role of caveolae in NO signalling is critical since endothelial NO synthase activity is blocked by binding to caveolin-1 and activation of NO synthase is associated with the release from the inhibitory clamp of caveolin-1 [152]. Besides NO, endothelial cells and cardiomyocytes communicate by ET-1, angiotensin II, prostaglandin, peptide growth factors, neuregulin (reviewed in [153]). While for normal modulation of contractile cells by endothelial cells, specific levels of such second messengers are optional, a significant

alteration of these levels may augment pathologies like myocardial infarction, ischaemia, hypertension, arrhythmias, congestive heart failure, and atherosclerosis [154-157].

In addition to gap and tight junctions, endothelial cells communicate with their neighbours through adherens junctions. In this type of junction their cytoplasmic surface is linked to the actin cytoskeleton. They are expressed as bands forming a circle around the cell (zonula adherens) or as loci of joints to the extracellular matrix (adhesion plaques). Similar cell junctions (fascia adherence) were found in cardiac myocyte which form on the surface of cardiac myocyte a ribbon-like structure not long enough to completely circle the cell. Adherent junctions contain cadherins,  $\alpha$ -catenin,  $\beta$ -catenin, p120 ( $\delta$ -catenin),  $\gamma$ -catenin (reviewed in [158]).

#### To Rejuvenate: Cross-Talk Between Organs and Organisms

Cross-talk between organs is well illustrated by the data in studies on protection of the brain or heart (damaged during a stroke or infarct correspondingly) by a remote preconditioning of the kidney or limbs [159-163]. The concept of remote preconditioning has evolved into 'remote conditioning', a term that encompasses a number of related endogenous cardioprotective strategies, applied to remote organs before (remote ischaemic preconditioning), during (preconditioning), or after (postconditioning) acute myocardial infarction [164,165]. Remote organ-heart neuronal and humoral communications can afford protection to the heart against stresses (i.g., by erythropoietin synthesised in kidney and liver [166,167] or adenosine, bradykinin, stromal derived factor- $1\alpha$  and others (for review see [165])). In contrast, considering pathophysiological aspects, heart failure is often accompanied by a number of comorbidities of the kidney [168,169], liver [170,171] or other organs [172,173], and thus adverse communication may be involved.

In the hierarchy of the biological communicational systems, the cross-talk between organisms joined together either artificially (such as in parabiosis [174]) or naturally (such as at pregnancy [175]) with at least partial unification of the blood systems, further demonstrate the potency of cross-talk between organs. Parabiosis is an example of a remote communication whereby humoral factors circulate and transfer from one parabiotic subject to another, i.e., when only one parabiotic partner was exposed to low pO<sub>2</sub>, erythropoiesis was observed in both partners [176] due to induced synthesis of erythropoietin in both. In general, the parabiotic interaction has common features to the junctional connection of two cells. Both interactions can result in functional changes in both partners thus playing an either deleterious or healing/rejuvenating role (See Figure 2). In recent studies, the regenerative (possibly rejuvenating) effect of parabiosis on, skeletal muscles [177], liver [178] and brain [179-181] has been demonstrated. The study on rejuvenation of the ageing heart by using the parabiotic model is most intriguing [182]. It was shown that after four

weeks of exposure to the circulation of young mice, the aged heart showed significantly regressed cardiac hypertrophy and molecular remodelling while growth factor 11 (GDF11) simulated the positive outcome reached by parabiosis. In the parabiotic model, the organisms do not communicate through large vessels, although they are united by small vessels, mostly capillaries (by the way, young capillaries have been considered a rejuvenation factor [183]) which were suggested to serve a root for trafficking of rejuvenating factors including GDF11 from young parabionts. Recombinant GDF11 was shown to induce inhibition of phenylephrine-mediated hypertrophy in cardiac myocytes supporting the idea that cardiac myocytes are primary targets for GDF11 [182]. In addition, GDF11 reversed impairments in aged muscle stem cells (satellite cells) [184] showing the high potential of this rejuvenating factor to regenerative therapy leading to improvements in cardiac performance [185]. However, due to methodological controversies [186] full experimental evidence supporting the role of GDF11 as a rejuvenating factor is currently incomplete. Similar to the many studies that have examined the multiple components of conditioned media for factors involved in stimulating stem cells, vascular or myocardial cells, or other cell types, communicating beneficial and adaptive intercellular signalling, such factors in parabiotic communication also remain a focus of research [187-189].

#### Conclusion

In the present review we have highlighted the importance of intercellular cross-talk mechanistic processes for homeostatic maintenance between differing neighbouring and distant cells during adaptation and survival as a tissue and organ. Such processes are highly diverse and are still being studied at a rudimentary level, mainly in experimental models and require considerable further research in order to fully determine their specific roles in normal physiology and pathology. Continued research in these basic processes will afford greater mechanistic insights into targeting disease aetiology and potential future therapeutic targets.

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#### References

- [1] Camelliti P, Borg TK, Kohl P. Structural and functional characterisation of cardiac fibroblasts. Cardiovasc Res 2005;65:40-51.
- [2] Kakkar R, Lee RT. Intramyocardial fibroblast myocyte communication . Circ Res 2010;106:47-57.
- [3] Johnstone RM, Adam M, Hammond JR, Orr L, Turbide C. Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes). J Biol Chem 1987;262:9412-20.
- [4] Valadi H, Ekstrom K, Bossios A, Sjostrand M, Lee JJ, Lotvall JO. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. Nat Cell Biol 2007;9:654-9.
- [5] Malik ZA, Kott KS, Poe AJ, Kuo T, Chen L, Ferrara KW et al. Cardiac myocyte exosomes: stability, HSP60, and proteomics. Am J Physiol Heart Circ Physiol 2013;304:H954-65.
- [6] Tominaga N, Kosaka N, Ono M, Katsuda T, Yoshioka Y, Tamura K et al. Brain metastatic cancer cells release microRNA-181c-containing extracellular vesicles capable of destructing blood-brain barrier. Nat Commun 2015;6:6716.
- [7] Court FA, Hendriks WT, MacGillavry HD, Alvarez J, van Minnen J. Schwann cell to axon transfer of ribosomes: toward a novel understanding of the role of glia in the nervous system. J Neurosci 2008;28:11024-9.
- [8] Twiss JL, Fainzilber M. Ribosomes in axons--scrounging from the neighbors? Trends Cell Biol 2009;19:236-43.
- [9] Sun D, Zhuang X, Zhang S, Deng ZB, Grizzle W, Miller D et al. Exosomes are endogenous nanoparticles that can deliver biological information between cells. Adv Drug Deliv Rev 2013;65:342-7.
- [10] Jaiswal N, Jaiswal RK, Tallant EA, Diz DI, Ferrario CM. Alterations in prostaglandin production in spontaneously hypertensive rat smooth muscle cells. Hypertension 1993;21:900-5.
- [11] Porto I, De Maria GL, Di Vito L, Camaioni C, Gustapane M, Biasucci LM. Microparticles in health and disease: small mediators, large role? Curr Vasc Pharmacol 2011;9:490-500.
- [12] Viera AJ, Mooberry M, Key NS. Microparticles in cardiovascular disease pathophysiology and outcomes. J Am Soc Hypertens 2012;6:243-52.
- [13] Arslan F, Lai RC, Smeets MB, Akeroyd L, Choo A, Aguor EN et al. Mesenchymal stem cell-derived exosomes increase ATP levels, decrease oxidative stress and activate PI3K/Akt pathway to enhance myocardial viability and prevent adverse remodeling after myocardial ischemia/reperfusion injury. Stem Cell Res 2013;10:301-12.
- [14] Barile L, Moccetti T, Marban E, Vassalli G. Roles of exosomes in cardioprotection. Eur Heart J 2016.
- [15] Desplantez T, Dupont E, Severs NJ, Weingart R. Gap junction channels and cardiac impulse propagation. J Membr Biol 2007;218:13-28.
- [16] Hoh JH, Lal R, John SA, Revel JP, Arnsdorf MF. Atomic force microscopy and dissection of gap junctions. Science 1991;253:1405-8.
- [17] Neijssen J, Herberts C, Drijfhout JW, Reits E, Janssen L, Neefjes J. Cross-presentation by intercellular peptide transfer through gap junctions. Nature 2005;434:83-8.
- [18] De Mello WC. Cell-to-cell diffusion of glucose in the mammalian heart is disrupted by high glucose. Implications for the diabetic heart. Exp Cell Res 2015;334:239-45.

- [19] Huan T, Rong J, Tanriverdi K, Meng Q, Bhattacharya A, McManus DD et al. Dissecting the roles of microRNAs in coronary heart disease via integrative genomic analyses. Arterioscler Thromb Vasc Biol 2015;35:1011-21.
- [20] De Mello WC. Effect of intracellular injection of calcium and strontium on cell communication in heart. J Physiol 1975;250:231-45.
- [21] Matsuda H, Kurata Y, Oka C, Matsuoka S, Noma A. Magnesium gating of cardiac gap junction channels. Prog Biophys Mol Biol 2010;103:102-10.
- [22] Dakhlallah D, Zhang J, Yu L, Marsh CB, Angelos MG, Khan M. MicroRNA-133a engineered mesenchymal stem cells augment cardiac function and cell survival in the infarct heart. J Cardiovasc Pharmacol 2015;65:241-51.
- [23] Icli B, Wara AK, Moslehi J, Sun X, Plovie E, Cahill M et al. MicroRNA-26a regulates pathological and physiological angiogenesis by targeting BMP/SMAD1 signaling. Circ Res 2013;113 :1231-41.
- [24] van Rooij E, Sutherland LB, Thatcher JE, DiMaio JM, Naseem RH, Marshall WS, et al. Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis. Proc Natl Acad Sci USA 2008;105:13027-32.
- [25] Rustom A, Saffrich R, Markovic I, Walther P, Gerdes HH. Nanotubular highways for intercellular organelle transport. Science 2004;303:1007-10.
- [26] Astanina K, Koch M, Jungst C, Zumbusch A, Kiemer AK. Lipid droplets as a novel cargo of tunnelling nanotubes in endothelial cells. Sci Rep 2015;5:11453.
- [27] Sisakhtnezhad S, Khosravi L. Emerging physiological and pathological implications of tunneling nanotubes formation between cells. Eur J Cell Biol 2015;94:429-43.
- [28] Sowinski S, Jolly C, Berninghausen O, Purbhoo MA, Chauveau A, Kohler K et al. Membrane nanotubes physically connect T cells over long distances presenting a novel route for HIV-1 transmission. Nat Cell Biol 2008;10:211-9.
- [29] Wang X, Veruki ML, Bukoreshtliev NV, Hartveit E, Gerdes HH. Animal cells connected by nanotubes can be electrically coupled through interposed gap-junction channels. Proc Natl Acad Sci USA 2010;107:17194-9.
- [30] Bukoreshtliev NV, Wang X, Hodneland E, Gurke S, Barroso JF, Gerdes HH. Selective block of tunneling nanotube (TNT) formation inhibits intercellular organelle transfer between PC12 cells. FEBS Lett 2009;583:1481-8.
- [31] Onfelt B, Nedvetzki S, Benninger RK, Purbhoo MA, Sowinski S, Hume AN et al. Structurally distinct membrane nanotubes between human macrophages support long-distance vesicular traffic or surfing of bacteria. J Immunol 2006;177:8476-83.
- [32] Plotnikov EY, Khryapenkova TG, Vasileva AK, Marey MV, Galkina SI, Isaev NK et al. Cell-to-cell cross-talk between mesenchymal stem cells and cardiomyocytes in co-culture. J Cell Mol Med 2008;12:1622-31.
- [33] Karlsson A, Karlsson R, Karlsson M, Cans AS, Stromberg A, Ryttsen F et al. Networks of nanotubes and containers. Nature 2001;409:150-2.
- [34] Mulcahy LA, Pink RC, Carter DR. Routes and mechanisms of extracellular vesicle uptake. J Extracell Vesicles 2014;3.
- [35] Thayanithy V, Babatunde V, Dickson EL, Wong P, Oh S, Ke X et al. Tumor exosomes induce tunneling nanotubes in lipid raft-enriched regions of human mesothelioma cells. Exp Cell Res 2014;323:178-88.

- [36] Pellman J, Zhang J, Sheikh F. Myocyte-fibroblast communication in cardiac fibrosis and arrhythmias: Mechanisms and model systems. J Mol Cell Cardiol 2016;94:22-31.
- [37] Lokar M, Kabaso D, Resnik N, Sepcic K, Kralj-Iglic V, Veranic P et al. The role of cholesterolsphingomyelin membrane nanodomains in the stability of intercellular membrane nanotubes. Int J Nanomedicine 2012;7:1891-902.
- [38] Gupta N, DeFranco AL. Lipid rafts and B cell signaling. Semin Cell Dev Biol 2007;18:616-26.
- [39] Mulcahy LA, Pink RC, Carter DR. Routes and mechanisms of extracellular vesicle uptake. J Extracell Vesicles 2014;3.
- [40] See Hoe LE, Schilling JM, Tarbit E, Kiessling CJ, Busija AR, Niesman IR et al. Sarcolemmal cholesterol and caveolin-3 dependence of cardiac function, ischemic tolerance, and opioidergic cardioprotection. Am J Physiol Heart Circ Physiol 2014;307:H895-903.
- [41] Paulus WJ, Tschope C. A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. J Am Coll Cardiol 2013;62:263-71.
- [42] Frangogiannis NG. The inflammatory response in myocardial injury, repair, and remodelling. Nat Rev Cardiol 2014;11:255-65.
- [43] Westermann D, Lindner D, Kasner M, Zietsch C, Savvatis K, Escher F et al. Cardiac inflammation contributes to changes in the extracellular matrix in patients with heart failure and normal ejection fraction. Circ Heart Fail 2011;4:44-52.
- [44] Norcross MA. A synaptic basis for T-lymphocyte activation. Ann Immunol (Paris) 1984;135D:113-34.
- [45] Kucik DF, Dustin ML, Miller JM, Brown EJ. Adhesion-activating phorbol ester increases the mobility of leukocyte integrin LFA-1 in cultured lymphocytes. J Clin Invest 1996;97:2139-44.
- [46] Sanchez-Madrid F, del Pozo MA. Leukocyte polarization in cell migration and immune interactions. EMBO J 1999;18:501-11.
- [47] Bromley SK, Burack WR, Johnson KG, Somersalo K, Sims TN, Sumen C et al. The immunological synapse. Annu Rev Immunol 2001;19:375-96.
- [48] Buck MD, O'Sullivan D, Klein Geltink RI, Curtis JD, Chang CH, Sanin DE et al. Mitochondrial Dynamics Controls T Cell Fate through Metabolic Programming. Cell 2016;166:63-76.
- [49] Timmerman I, Daniel AE, Kroon J, van Buul JD. Leukocytes Crossing the Endothelium: A Matter of Communication. Int Rev Cell Mol Biol 2016;322:281-329.
- [50] Butcher EC. Leukocyte-endothelial cell recognition: three (or more) steps to specificity and diversity. Cell 1991;67:1033-6.
- [51] Nourshargh S, Hordijk PL, Sixt M. Breaching multiple barriers: leukocyte motility through venular walls and the interstitium. Nat Rev Mol Cell Biol 2010;11:366-78.
- [52] Butoi E, Gan AM, Tucureanu MM, Stan D, Macarie RD, Constantinescu C et al. Cross-talk between macrophages and smooth muscle cells impairs collagen and metalloprotease synthesis and promotes angiogenesis. Biochim Biophys Acta 2016;1863:1568-78.
- [53] Gao Y, Chen T, Raj JU. Endothelial and Smooth Muscle Cell Interactions in the Pathobiology of Pulmonary Hypertension. Am J Respir Cell Mol Biol 2016;54:451-60.
- [54] Balcells M, Martorell J, Olive C, Santacana M, Chitalia V, Cardoso AA et al. Smooth muscle cells orchestrate the endothelial cell response to flow and injury. Circulation 2010;121 :2192-9.

- [55] Billaud M, Lohman AW, Johnstone SR, Biwer LA, Mutchler S, Isakson BE. Regulation of cellular communication by signaling microdomains in the blood vessel wall. Pharmacol Rev 2014;66:513-69.
- [56] Brucher BL, Jamall IS . Epistemology of the origin of cancer: a new paradigm. BMC Cancer 2014;14:331.
- [57] Becker BF, Chappell D, Bruegger D, Annecke T, Jacob M. Therapeutic strategies targeting the endothelial glycocalyx: acute deficits, but great potential. Cardiovasc Res 2010;87:300-10.
- [58] Brucher BL, Jamall IS. Cell-cell communication in the tumor microenvironment, carcinogenesis, and anticancer treatment. Cell Physiol Biochem 2014;34:213-43.
- [59] Zhang P, Su J, Mende U. Cross talk between cardiac myocytes and fibroblasts: from multiscale investigative approaches to mechanisms and functional consequences. Am J Physiol Heart Circ Physiol 2012;303:H1385-96.
- [60] Box C, Rogers SJ, Mendiola M, Eccles SA. Tumour-microenvironmental interactions: paths to progression and targets for treatment. Semin Cancer Biol 2010;20:128-38.
- [61] Komuro T. Re-evaluation of fibroblasts and fibroblast-like cells. Anat Embryol (Berl) 1990;182:103-12.
- [62] Schmitt-Graff A, Desmouliere A, Gabbiani G. Heterogeneity of myofibroblast phenotypic features: an example of fibroblastic cell plasticity. Virchows Arch 1994;425:3-24.
- [63] Prabhu SD, Frangogiannis NG. The Biological Basis for Cardiac Repair After Myocardial Infarction: From Inflammation to Fibrosis. Circ Res 2016;119:91-112.
- [64] Tan AS, Baty JW, Dong LF, Bezawork-Geleta A, Endaya B, Goodwin J et al. Mitochondrial genome acquisition restores respiratory function and tumorigenic potential of cancer cells without mitochondrial DNA. Cell Metab 2015;21:81-94.
- [65] Yousefi S, Gold JA, Andina N, Lee JJ, Kelly AM, Kozlowski E et al. Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. Nat Med 2008;14:949-53.
- [66] McCully JD, Levitsky S, Del Nido PJ, Cowan DB. Mitochondrial transplantation for therapeutic use. Clin Transl Med 2016;5:16.
- [67] Miranda CH, Borges Mde C, Matsuno AK, Vilar FC, Gali LG, Volpe GJ et al. Evaluation of cardiac involvement during dengue viral infection. Clin Infect Dis 2013;57:812-9.
- [68] Kushawaha A, Jadonath S, Mobarakai N. West Nile virus myocarditis causing a fatal arrhythmia: a case report. Cases J 2009;2:7147.
- [69] Obeyesekere I, Hermon Y. Myocarditis and cardiomyopathy after arbovirus infections (dengue and chikungunya fever). Br Heart J 1972;34:821-7.
- [70] Vidricaire G, Tremblay MJ. A clathrin, caveolae, and dynamin-independent endocytic pathway requiring free membrane cholesterol drives HIV-1 internalization and infection in polarized trophoblastic cells. J Mol Biol 2007;368:1267-83.
- [71] Delorme-Axford E, Sadovsky Y, Coyne CB. The Placenta as a Barrier to Viral Infections. Annu Rev Virol 2014;1:133-46.
- [72] Naveiras O, Daley GQ. Stem cells and their niche: a matter of fate. Cell Mol Life Sci 2006;63:760-6.
- [73] Ilmer M, Vykoukal J, Recio Boiles A, Coleman M, Alt E. Two sides of the same coin: stem cells in cancer and regenerative medicine. FASEB J 2014;28:2748-61.
- [74] Sanchez-Aguilera A, Mendez-Ferrer S. The hematopoietic stem-cell niche in health and leukemia. Cell

Mol Life Sci 2016.

- [75] Hodgkinson CP, Bareja A, Gomez JA, Dzau VJ. Emerging Concepts in Paracrine Mechanisms in Regenerative Cardiovascular Medicine and Biology. Circ Res 2016;118:95-107.
- [76] Kourembanas S. Exosomes: vehicles of intercellular signaling, biomarkers, and vectors of cell therapy. Annu Rev Physiol 2015;77:13-27.
- [77] De Jong OG, Van Balkom BW, Schiffelers RM, Bouten CV, Verhaar MC. Extracellular vesicles: potential roles in regenerative medicine. Front Immunol 2014;5:608.
- [78] Lai RC, Yeo RW, Lim SK. Mesenchymal stem cell exosomes. Semin Cell Dev Biol 2015;40:82-8.
- [79] Menasche P, Vanneaux V. Stem cells for the treatment of heart failure. Curr Res Transl Med 2016;64:97-106.
- [80] Colombo M, Raposo G, Thery C. Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. Annu Rev Cell Dev Biol 2014;30:255-89.
- [81] Freund D, Bauer N, Boxberger S, Feldmann S, Streller U, Ehninger G et al. Polarization of human hematopoietic progenitors during contact with multipotent mesenchymal stromal cells: effects on proliferation and clonogenicity. Stem Cells Dev 2006;15:815-29.
- [82] Koyanagi M, Brandes RP, Haendeler J, Zeiher AM, Dimmeler S. Cell-to-cell connection of endothelial progenitor cells with cardiac myocytes by nanotubes: a novel mechanism for cell fate changes? Circ Res 2005;96:1039-41.
- [83] Plotnikov EY, Khryapenkova TG, Galkina SI, Sukhikh GT, Zorov DB. Cytoplasm and organelle transfer between mesenchymal multipotent stromal cells and renal tubular cells in co-culture. Exp Cell Res 2010;316:2447-55.
- [84] Rogers RS, Bhattacharya J. When cells become organelle donors. Physiology (Bethesda) 2013;28:414-22.
- [85] Islam MN, Das SR, Emin MT, Wei M, Sun L, Westphalen K et al. Mitochondrial transfer from bonemarrow-derived stromal cells to pulmonary alveoli protects against acute lung injury. Nat Med 2012;18:759-65.
- [86] Ahmad T, Mukherjee S, Pattnaik B, Kumar M, Singh S, Kumar M et al. Miro1 regulates intercellular mitochondrial transport & enhances mesenchymal stem cell rescue efficacy. EMBO J 2014;33:994-1010.
- [87] Li X, Zhang Y, Yeung SC, Liang Y, Liang X, Ding Y et al. Mitochondrial transfer of induced pluripotent stem cell-derived mesenchymal stem cells to airway epithelial cells attenuates cigarette smoke-induced damage. Am J Respir Cell Mol Biol 2014;51:455-65.
- [88] Otsu K, Das S, Houser SD, Quadri SK, Bhattacharya S, Bhattacharya J. Concentration-dependent inhibition of angiogenesis by mesenchymal stem cells. Blood 2009;113:4197-205.
- [89] Vallabhaneni KC, Haller H, Dumler I. Vascular smooth muscle cells initiate proliferation of mesenchymal stem cells by mitochondrial transfer via tunneling nanotubes. Stem Cells Dev 2012;21:3104-13.
- [90] Acquistapace A, Bru T, Lesault PF, Figeac F, Coudert AE, le Coz O et al. Human mesenchymal stem cells reprogram adult cardiomyocytes toward a progenitor-like state through partial cell fusion and mitochondria transfer. Stem Cells 2011;29:812-24.
- [91] Vasquez C, Benamer N, Morley GE. The cardiac fibroblast: functional and electrophysiological considerations in healthy and diseased hearts . J Cardiovasc Pharmacol 2011;57:380-8.

- [92] Swynghedauw B. Molecular mechanisms of myocardial remodeling. Physiol Rev 1999;79:215-62.
- [93] Lakatta EG, Sollott SJ. Perspectives on mammalian cardiovascular aging: humans to molecules. Comp Biochem Physiol A Mol Integr Physiol 2002;132:699-721.
- [94] Baudino TA, Carver W, Giles W, Borg TK. Cardiac fibroblasts: friend or foe? Am J Physiol Heart Circ Physiol 2006;291:H1015-26.
- [95] Porter KE, Turner NA. Cardiac fibroblasts: at the heart of myocardial remodeling. Pharmacol Ther 2009;123:255-78.
- [96] Ottaviano FG, Yee KO. Communication signals between cardiac fibroblasts and cardiac myocytes. J Cardiovasc Pharmacol 2011;57:513-21.
- [97] Chen MM, Lam A, Abraham JA, Schreiner GF, Joly AH. CTGF expression is induced by TGF- beta in cardiac fibroblasts and cardiac myocytes: a potential role in heart fibrosis. J Mol Cell Cardiol 2000;32:1805-19.
- [98] Sanada S, Hakuno D, Higgins LJ, Schreiter ER, McKenzie AN, Lee RT. IL-33 and ST2 comprise a critical biomechanically induced and cardioprotective signaling system. J Clin Invest 2007;117:1538-49.
- [99] Seki K, Sanada S, Kudinova AY, Steinhauser ML, Handa V, Gannon J et al. Interleukin-33 prevents apoptosis and improves survival after experimental myocardial infarction through ST2 signaling. Circ Heart Fail 2009;2:684-91.
- [100] Merle PL, Feige JJ, Verdetti J. Basic fibroblast growth factor activates calcium channels in neonatal rat cardiomyocytes. J Biol Chem 1995;270:17361-7.
- [101] Doble BW, Chen Y, Bosc DG, Litchfield DW, Kardami E. Fibroblast growth factor-2 decreases metabolic coupling and stimulates phosphorylation as well as masking of connexin43 epitopes in cardiac myocytes. Circ Res 1996;79:647-58.
- [102] Pedrotty DM, Klinger RY, Badie N, Hinds S, Kardashian A, Bursac N. Structural coupling of cardiomyocytes and noncardiomyocytes: quantitative comparisons using a novel micropatterned cell pair assay. Am J Physiol Heart Circ Physiol 2008;295:H390-400.
- [103] Nishida M, Sato Y, Uemura A, Narita Y, Tozaki-Saitoh H, Nakaya M et al. P2Y6 receptor-Galpha12/13 signalling in cardiomyocytes triggers pressure overload-induced cardiac fibrosis. EMBO J 2008;27:3104-15.
- [104] Shestopalov VI, Panchin Y. Pannexins and gap junction protein diversity. Cell Mol Life Sci 2008;65:376-94.
- [105] Rohr S, Scholly DM, Kleber AG. Patterned growth of neonatal rat heart cells in culture. Morphological and electrophysiological characterization. Circ Res 1991;68:114-30.
- [106] Askar SF, Bingen BO, Swildens J, Ypey DL, van der Laarse A, Atsma DE et al. Connexin43 silencing in myofibroblasts prevents arrhythmias in myocardial cultures: role of maximal diastolic potential. Cardiovasc Res 2012;93:434-44.
- [107] Xie Y, Garfinkel A, Weiss JN, Qu Z. Cardiac alternans induced by fibroblast-myocyte coupling: mechanistic insights from computational models. Am J Physiol Heart Circ Physiol 2009;297:H775-84.
- [108] Hasan W. Autonomic cardiac innervation: development and adult plasticity. Organogenesis 2013;9:176-93.
- [109] del Zoppo GJ. The neurovascular unit, matrix proteases, and innate inflammation. Ann N Y Acad Sci 2010;1207:46-9.

- [110] Silachev DN, Plotnikov EY, Babenko VA, Savchenko ES, Zorova LD, Pevzner IB et al. Protection of Neurovascular Unit Cells with Lithium Chloride and Sodium Valproate Prevents Brain Damage in Neonatal Ischemia/Hypoxia. Bull Exp Biol Med 2016;160:313-8.
- [111] Agnati LF, Fuxe K. Volume transmission as a key feature of information handling in the central nervous system possible new interpretative value of the Turing's B-type machine. Prog Brain Res 2000;125:3-19.
- [112] Rosenberg PA, Aizenman E. Hundred-fold increase in neuronal vulnerability to glutamate toxicity in astrocyte-poor cultures of rat cerebral cortex. Neurosci Lett 1989;103 :162-8.
- [113] Haidet-Phillips AM, Hester ME, Miranda CJ, Meyer K, Braun L, Frakes A et al. Astrocytes from familial and sporadic ALS patients are toxic to motor neurons. Nat Biotechnol 2011;29:824-8.
- [114] Davis CH, Kim KY, Bushong EA, Mills EA, Boassa D, Shih T et al. Transcellular degradation of axonal mitochondria. Proc Natl Acad Sci USA 2014;111:9633-8.
- [115] Hayakawa K, Esposito E, Wang X, Terasaki Y, Liu Y, Xing C et al. Transfer of mitochondria from astrocytes to neurons after stroke. Nature 2016;535:551-5.
- [116] Cowan DB, Yao R, Akurathi V, Snay ER, Thedsanamoorthy JK, Zurakowski D et al. Intracoronary Delivery of Mitochondria to the Ischemic Heart for Cardioprotection. PLoS One 2016;11:e0160889.
- [117] Masuzawa A, Black KM, Pacak CA, Ericsson M, Barnett RJ, Drumm C et al. Transplantation of autologously derived mitochondria protects the heart from ischemia-reperfusion injury. Am J Physiol Heart Circ Physiol 2013;304:H966-82.
- [118] Falchi AM, Sogos V, Saba F, Piras M, Congiu T, Piludu M. Astrocytes shed large membrane vesicles that contain mitochondria, lipid droplets and ATP. Histochem Cell Biol 2013;139:221-31.
- [119] Langer J, Stephan J, Theis M, Rose CR. Gap junctions mediate intercellular spread of sodium between hippocampal astrocytes in situ. Glia 2012;60:239-52.
- [120] Rose CR, Chatton JY. Astrocyte sodium signaling and neuro-metabolic coupling in the brain. Neuroscience 2016;323:121-34.
- [121] Dermietzel R, Traub O, Hwang TK, Beyer E, Bennett MV, Spray DC et al. Differential expression of three gap junction proteins in developing and mature brain tissues. Proc Natl Acad Sci USA 1989;86:10148-52.
- [122] Nagy JI, Patel D, Ochalski PA, Stelmack GL. Connexin30 in rodent, cat and human brain: selective expression in gray matter astrocytes, co-localization with connexin43 at gap junctions and late developmental appearance. Neuroscience 1999;88:447-68.
- [123] Dermietzel R, Gao Y, Scemes E, Vieira D, Urban M, Kremer M et al. Connexin43 null mice reveal that astrocytes express multiple connexins. Brain Res Brain Res Rev 2000;32:45-56.
- [124] Orellana JA, Stehberg J. Hemichannels: new roles in astroglial function. Front Physiol 2014;5:193.
- [125] Fu J, Lee K, Chuang PY, Liu Z, He JC. Glomerular endothelial cell injury and cross talk in diabetic kidney disease. Am J Physiol Renal Physiol 2015;308:F287-97.
- [126] Weil EJ, Lemley KV, Mason CC, Yee B, Jones LI, Blouch K et al. Podocyte detachment and reduced glomerular capillary endothelial fenestration promote kidney disease in type 2 diabetic nephropathy. Kidney Int 2012;82:1010-7.
- [127] Kuwabara A, Satoh M, Tomita N, Sasaki T, Kashihara N. Deterioration of glomerular endothelial surface layer induced by oxidative stress is implicated in altered permeability of macromolecules in Zucker fatty rats. Diabetologia 2010;53:2056-65.

- [128] Cha DR, Kang YS, Han SY, Jee YH, Han KH, Han JY et al. Vascular endothelial growth factor is increased during early stage of diabetic nephropathy in type II diabetic rats. J Endocrinol 2004;183:183-94.
- [129] Chen S, Kasama Y, Lee JS, Jim B, Marin M, Ziyadeh FN. Podocyte-derived vascular endothelial growth factor mediates the stimulation of alpha3(IV) collagen production by transforming growth factor-beta1 in mouse podocytes. Diabetes 2004;53:2939-49.
- [130] Kedzierski RM, Yanagisawa M. Endothelin system: the double-edged sword in health and disease. Annu Rev Pharmacol Toxicol 2001;41:851-76.
- [131] Woolf AS, Gnudi L, Long DA. Roles of angiopoietins in kidney development and disease. J Am Soc Nephrol 2009;20:239-44.
- [132] Haraldsson BS. The endothelium as part of the integrative glomerular barrier complex. Kidney Int 2014;85:8-11.
- [133] Xu C, Chang A, Hack BK, Eadon MT, Alper SL, Cunningham PN. TNF-mediated damage to glomerular endothelium is an important determinant of acute kidney injury in sepsis. Kidney Int 2014;85:72-81.
- [134] Yuen DA, Stead BE, Zhang Y, White KE, Kabir MG, Thai K et al. eNOS deficiency predisposes podocytes to injury in diabetes. J Am Soc Nephrol 2012;23:1810-23.
- [135] Cheng H, Wang S, Jo YI, Hao CM, Zhang M, Fan X et al. Overexpression of cyclooxygenase-2 predisposes to podocyte injury. J Am Soc Nephrol 2007;18:551-9.
- [136] Hergenreider E, Heydt S, Treguer K, Boettger T, Horrevoets AJ, Zeiher AM et al. Atheroprotective communication between endothelial cells and smooth muscle cells through miRNAs. Nat Cell Biol 2012;14:249-56.
- [137] Wildhirt SM, Dudek RR, Suzuki H, Bing RJ. Involvement of inducible nitric oxide synthase in the inflammatory process of myocardial infarction. Int J Cardiol 1995;50:253-61.
- [138] Balligand JL, Kobzik L, Han X, Kaye DM, Belhassen L, O'Hara DS et al. Nitric oxide-dependent parasympathetic signaling is due to activation of constitutive endothelial (type III) nitric oxide synthase in cardiac myocytes. J Biol Chem 1995;270:14582-6.
- [139] Qian J, Zhang Q, Church JE, Stepp DW, Rudic RD, Fulton DJ. Role of local production of endotheliumderived nitric oxide on cGMP signaling and S-nitrosylation. Am J Physiol Heart Circ Physiol 2010;298:H112-8.
- [140] Hobbs AJ. Soluble guanylate cyclase: the forgotten sibling. Trends Pharmacol Sci 1997;18:484-91.
- [141] Abi-Gerges N, Szabo G, Otero AS, Fischmeister R, Mery PF. NO donors potentiate the beta-adrenergic stimulation of I(Ca,L) and the muscarinic activation of I(K,ACh) in rat cardiac myocytes. J Physiol 2002;540:411-24.
- [142] Petroff MG, Kim SH, Pepe S, Dessy C, Marban E, Balligand JL et al. Endogenous nitric oxide mechanisms mediate the stretch dependence of Ca2+ release in cardiomyocytes. Nat Cell Biol 2001;3:867-73.
- [143] Maguy A, Hebert TE, Nattel S. Involvement of lipid rafts and caveolae in cardiac ion channel function. Cardiovasc Res 2006;69:798-807.
- [144] Fridolfsson HN, Kawaraguchi Y, Ali SS, Panneerselvam M, Niesman IR, Finley JC et al. Mitochondrialocalized caveolin in adaptation to cellular stress and injury. FASEB J 2012;26:4637-49.
- [145] Wang J, Schilling JM, Niesman IR, Headrick JP, Finley JC, Kwan E et al. Cardioprotective trafficking of caveolin to mitochondria is Gi-protein dependent. Anesthesiology 2014;121:538-48.

- [146] Li WP, Liu P, Pilcher BK, Anderson RG. Cell-specific targeting of caveolin-1 to caveolae, secretory vesicles, cytoplasm or mitochondria. J Cell Sci 2001;114:1397-408.
- [147] Fridolfsson HN, Kawaraguchi Y, Ali SS, Panneerselvam M, Niesman IR, Finley JC et al. Mitochondrialocalized caveolin in adaptation to cellular stress and injury. FASEB J 2012;26:4637-49.
- [148] Patel HH, Murray F, Insel PA. Caveolae as organizers of pharmacologically relevant signal transduction molecules. Annu Rev Pharmacol Toxicol 2008;48:359-91.
- [149] Tsutsumi YM, Horikawa YT, Jennings MM, Kidd MW, Niesman IR, Yokoyama U et al. Cardiac-specific overexpression of caveolin-3 induces endogenous cardiac protection by mimicking ischemic preconditioning. Circulation 2008;118:1979-88.
- [150] Juhaszova M, Zorov DB, Kim SH, Pepe S, Fu Q, Fishbein KW et al. Glycogen synthase kinase-3beta mediates convergence of protection signaling to inhibit the mitochondrial permeability transition pore. J Clin Invest 2004;113:1535-49.
- [151] Rodriguez-Sinovas A, Boengler K, Cabestrero A, Gres P, Morente M, Ruiz-Meana M et al. Translocation of connexin 43 to the inner mitochondrial membrane of cardiomyocytes through the heat shock protein 90-dependent TOM pathway and its importance for cardioprotection. Circ Res 2006;99:93-101.
- [152] Garcia-Cardena G, Martasek P, Masters BS, Skidd PM, Couet J, Li S et al. Dissecting the interaction between nitric oxide synthase (NOS) and caveolin. Functional significance of the nos caveolin binding domain in vivo. J Biol Chem 1997;272:25437-40.
- [153] Noireaud J, Andriantsitohaina R. Recent insights in the paracrine modulation of cardiomyocyte contractility by cardiac endothelial cells. Biomed Res Int 2014;2014:923805.
- [154] Rakhit A, Maguire CT, Wakimoto H, Gehrmann J, Li GK, Kelly RA et al. In vivo electrophysiologic studies in endothelial nitric oxide synthase (eNOS)-deficient mice. J Cardiovasc Electrophysiol 2001;12:1295-301.
- [155] Kuruvilla L, Kartha CC. Molecular mechanisms in endothelial regulation of cardiac function. Mol Cell Biochem 2003;253:113-23.
- [156] Carnicer R, Crabtree MJ, Sivakumaran V, Casadei B, Kass DA. Nitric oxide synthases in heart failure. Antioxid Redox Signal 2013;18:1078-99.
- [157] Drawnel FM, Archer CR, Roderick HL. The role of the paracrine/autocrine mediator endothelin-1 in regulation of cardiac contractility and growth. Br J Pharmacol 2013;168 :296-317.
- [158] Bazzoni G, Dejana E. Endothelial cell-to-cell junctions: molecular organization and role in vascular homeostasis. Physiol Rev 2004;84:869-901.
- [159] McClanahan B, Nao BS, Wolke LJ, Martin BJ, Metz TE, and Gallagher KP. Brief renal occlusion and reperfusion reduces myocardial infarct size in rabbits. FASEB J. 7.
- [160] Tapuria N, Kumar Y, Habib MM, Abu Amara M, Seifalian AM, Davidson BR. Remote ischemic preconditioning: a novel protective method from ischemia reperfusion injury--a review. J Surg Res 2008;150:304-30.
- [161] Silachev DN, Isaev NK, Pevzner IB, Zorova LD, Stelmashook EV, Novikova SV et al. The mitochondria-targeted antioxidants and remote kidney preconditioning ameliorate brain damage through kidney-to-brain cross-talk. PLoS One 2012;7:e51553.
- [162] Liu M, Liang Y, Chigurupati S, Lathia JD, Pletnikov M, Sun Z et al. Acute kidney injury leads to inflammation and functional changes in the brain. J Am Soc Nephrol 2008;19:1360-70.
- [163] Lu R, Kiernan MC, Murray A, Rosner MH, Ronco C. Kidney-brain crosstalk in the acute and chronic

setting. Nat Rev Nephrol 2015;11:707-19.

- [164] Hausenloy DJ,Yellon DM. Ischaemic conditioning and reperfusion injury. Nat Rev Cardiol 2016;13:193-209.
- [165] Heusch G, Botker HE, Przyklenk K, Redington A, Yellon D. Remote ischemic conditioning. J Am Coll Cardiol 2015;65:177-95.
- [166] Moon C, Krawczyk M, Ahn D, Ahmet I, Paik D, Lakatta EG et al. Erythropoietin reduces myocardial infarction and left ventricular functional decline after coronary artery ligation in rats. Proc Natl Acad Sci USA 2003;100:11612-7.
- [167] Sanchis-Gomar F, Garcia-Gimenez JL, Pareja-Galeano H, Romagnoli M, Perez-Quilis C, Lippi G. Erythropoietin and the heart: physiological effects and the therapeutic perspective. Int J Cardiol 2014;171:116-25.
- [168] Segall L, Nistor I, Covic A. Heart failure in patients with chronic kidney disease: a systematic integrative review. Biomed Res Int 2014;2014:937398.
- [169] Schefold JC, Filippatos G, Hasenfuss G, Anker SD, von Haehling S. Heart failure and kidney dysfunction: epidemiology, mechanisms and management. Nat Rev Nephrol 2016;12:610-23.
- [170] Lee SS. Cardiac abnormalities in liver cirrhosis. West J Med 1989;151:530-5.
- [171] Ruiz-del-Arbol L, Serradilla R. Cirrhotic cardiomyopathy. World J Gastroenterol 2015;21:11502-21.
- [172] Jain S, Goldstein DS. Cardiovascular dysautonomia in Parkinson disease: from pathophysiology to pathogenesis. Neurobiol Dis 2012;46:572-80.
- [173] Zhang MH. Rhabdomyolosis and its pathogenesis. World J Emerg Med 2012;3: 11-5.
- [174] Hill RT. Blood exchange and hormonic reactions in parabiotic rats. Exptl. Zool. 1932;63:203-234.
- [175] Popkov VA, Jankauskas SS, Silachev DN, Zorova LD, Pevzner IB, Plotnikov EY et al. Molecular and cellular interactions between mother and fetus. Pregnancy as rejuvenating factor. Biochemistry (Mosc) 2016.
- [176] Reissmann KR. Studies on the mechanism of erythropoietic stimulation in parabiotic rats during hypoxia. Blood 1950;5:372-80.
- [177] Conboy IM, Rando TA. Aging, stem cells and tissue regeneration: lessons from muscle. Cell Cycle 2005;4:407-10.
- [178] Conboy IM, Conboy MJ, Wagers AJ, Girma ER, Weissman IL, Rando TA. Rejuvenation of aged progenitor cells by exposure to a young systemic environment. Nature 2005;433:760-4.
- [179] Villeda SA, Luo J, Mosher KI, Zou B, Britschgi M, Bieri G et al. The ageing systemic milieu negatively regulates neurogenesis and cognitive function. Nature 2011;477:90-4.
- [180] Katsimpardi L, Litterman NK, Schein PA, Miller CM, Loffredo FS, Wojtkiewicz GR et al. Vascular and neurogenic rejuvenation of the aging mouse brain by young systemic factors. Science 2014;344:630-4.
- [181] Villeda SA, Plambeck KE, Middeldorp J, Castellano JM, Mosher KI, Luo J et al. Young blood reverses age-related impairments in cognitive function and synaptic plasticity in mice. Nat Med 2014;20:659-63.
- [182] Loffredo FS, Steinhauser ML, Jay SM, Gannon J, Pancoast JR, Yalamanchi P et al. Growth differentiation factor 11 is a circulating factor that reverses age-related cardiac hypertrophy. Cell 2013;153:828-39.
- [183] Almaca J, Molina J, Arrojo E Drigo R, Abdulreda MH, Jeon WB, Berggren PO et al. Young capillary

vessels rejuvenate aged pancreatic islets. Proc Natl Acad Sci USA 2014;111:17612-7.

- [184] Sinha M, Jang YC, Oh J, Khong D, Wu EY, Manohar R et al. Restoring systemic GDF11 levels reverses age-related dysfunction in mouse skeletal muscle. Science 2014;344:649-52.
- [185] Olson KA, Beatty AL, Heidecker B, Regan MC, Brody EN, Foreman T et al. Association of growth differentiation factor 11/8, putative anti-ageing factor, with cardiovascular outcomes and overall mortality in humans: analysis of the Heart and Soul and HUNT3 cohorts. Eur Heart J 2015;36:3426-34.
- [186] Smith SC, Zhang X, Zhang X, Gross P, Starosta T, Mohsin S et al. GDF11 does not rescue aging-related pathological hypertrophy. Circ Res 2015;117:926-32.
- [187] Dai W, Hale SL, Kloner RA. Role of a paracrine action of mesenchymal stem cells in the improvement of left ventricular function after coronary artery occlusion in rats. Regen Med 2007;2:63-8.
- [188] Rochette L, Zeller M, Cottin Y, Vergely C. Growth and differentiation factor 11 (GDF11): Functions in the regulation of erythropoiesis and cardiac regeneration. Pharmacol Ther 2015;156:26-33.
- [189] Rochette L, Vergely C. "Pro-youthful" factors in the "labyrinth" of cardiac rejuvenation. Exp Gerontol 2016;83:1-5.

Figures legends

Figure 1. Cardiac myocyte–fibroblast (left) and cardiac myocyte-stem cell (right) interaction in the heart. This includes electrical communication (through: 1, TNTs and 2, gap/adherence junctions); biochemical communication (through 3, EVs) and biomechanical communication (through 4, extracellular matrix). Some details can be found in [36]. While heart hypertrophy, arrhythmias and tissue fibrosis may result from interaction of cardiac myocyte with fibroblast, the interaction with stem cell can yield the healing effect through restoration of cellular bioenergetics and normalisation of electrical communication of cardiac myocytes along the tissue.

Figure 2. Parabiotic (left) and paracytotic (right) interactions. First is organised by unified circulation and second is organised by TNTs connecting neighbouring cells.



